



A review on hepatoprotective activity of Indian traditional plant

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Abstract

The liver is an essential organ that aids in the metabolism and elimination of xenobiotics from the body. Liver damage or liver dysfunction, is a serious health issue that concerns not only doctors, but also the pharmaceutical companies and drug regulatory bodies. The effects of numerous hazardous substances (particular antibiotics, chemotherapeutics, carbon tetrachloride (CCl₄), thioacetamide (TAA), and microorganisms on liver cells have been extensively researched. Synthetic drugs used to treat liver abnormalities in this condition can also harm the liver in other ways. Synthetic drugs used to treat liver abnormalities in this condition can also harm the liver in other ways. As a result, herbal medications are becoming increasingly popular, and their use is becoming more prevalent. The use of medicinal plants in the treatment of liver illness has a long history. On the market, there are a variety of herbal preparations. The goal of this review is to gather information on prospective phytochemicals from medicinal plants that have been investigated in modern scientific hepatotoxicity models.

Keywords: hepatoprotective, plant, liver

Introduction

The liver is one of the human body's most important organs, as it is the primary place for regulating metabolism and excretion. The liver is the body's second largest organ, weighing roughly 1.5 kg in adults, or 2% of total body weight [1]. It is responsible for the generation and excretion of bile, as well as the excretion of bilirubin, cholesterol, hormones, medications, proteins, and the metabolism of fats and carbohydrates. The liver's digestive fluid plays an important function in digesting, among other things [2]. The liver's physiological activity produces highly reactive free radicals with different molecular species in which an atomic constituent's unpaired valence electron does not contribute to binding inside free radical molecules. They are highly reactive due to the presence of free electrons from reactive oxygen species (ROS) [3]. Aging, energy production, the immune system, metabolism, toxins, and medicines are all primary sources of free radical generation in the body. Through covalent connections, free radicals attach to membrane lipids and cause lipid peroxidation [4]. Many chemicals, such as some chemotherapy medications, carbon tetrachloride, thioacetamide, and others, as well as persistent alcohol intake and microorganisms, cause damage to liver cells. Increased peroxidation of super molecules during alcohol metabolism may contribute to the development of liver disease [5]. Several earlier studies have found that liver damage increases the production of extracellular matrix proteins by liver stellate cells (ECM). An imbalance in the synthesis and usage of ECM results in an excessive accumulation of ECM, which leads to liver fibrosis [6]. Cirrhosis is an advanced stage of liver fibrosis with normal

vascular structure and disrupted parenchyma, which can be caused by liver fibrosis. Several investigations have demonstrated, however, that liver fibrosis and even cirrhosis are pathologically reversible and can be prevented early on. [7].

For decades, natural products have played an important role in healthcare. Plants have been a source of the chemical, functioning as full-fledged medications or important elements in formulations comprising synthetic drugs, and are often distinct sources of natural medicines [8]. The plant species chosen is a critical aspect in the research's possible success. Certain clues are revealed through random selection, and focused collection based on chemotaxonomic correlations and ethnomedical information gained from traditional medicine are more likely to yield pharmacologically active substances [9].

Despite advancements in modern medicine, herbal remedies continue to be in high demand. Before they may be validated for disease treatment, effective and potent herbal medicines must be evaluated using standard scientific methodologies [10].

Herbal medications are more extensively utilised as hepatoprotective treatments than allopathic medicines because they are less expensive, have a higher cultural acceptability, are more compatible with the human body, and produce minimal side effects. These natural medications have demonstrated the ability to preserve normal and useful liver stats while causing fewer adverse effects [11].

Causes of Hepatotoxicity

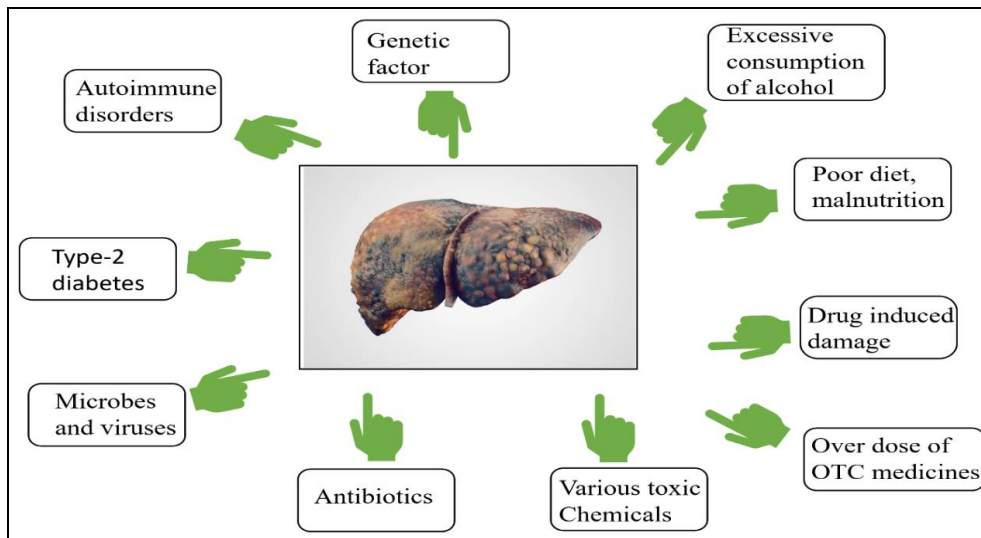


Fig 1: Common Causes of Hepatotoxicity

Mechanism of Hepatotoxicity

The bioactivation of medicines into chemically reactive metabolites, which can interact with cellular macromolecules like proteins, lipids, and nucleic acids, causing protein malfunction, lipid peroxidation, DNA damage, and stress oxidative stress, causes liver damage.^[12]

Furthermore, these reactive metabolites can cause ionic gradients and intracellular calcium reserves to be disrupted, resulting in mitochondrial malfunction and energy production loss.

Its malfunction causes an excess of oxidants, which damage liver cells^[13]. Oxidative stress is also caused by the activation with cytochrome P-450 system enzymes, such as CYP2E1. Bile acid builds up inside the liver due to damage to hepatocytes and bile duct cells. This causes the liver to be damaged even more. This change in cell function can result in cell death^[14].

In addition to triggering innate and adaptive immune responses, liver cell malfunction and death can also induce

immunological responses. Hepatocyte stress and damage trigger the release of signals that activate other cells, particularly those of the innate immune system, such as Kupffer cells (KC), or natural killer cells (NK)^[15]. These cells reduced the progress of liver injury by secreting chemokines and generating pro-inflammatory mediators that attracted more inflammatory cells to the liver. Several inflammatory cytokines released during liver injury, such as tumour necrosis factor (TNF)- α interferon (IFN)- γ , and interleukin (IL) - 1β , have been implicated in the promotion of tissue destruction.^[16]

Innate immune cells, on the other hand, are the main source of IL-10, IL-6, and certain prostaglandins, all of which have been demonstrated to have hepatoprotective properties. As a result, an individual's sensitivity and response to liver damage is determined by the delicate balance of inflammatory and hepatoprotective mediators released after activation of the innate immune system^[17].

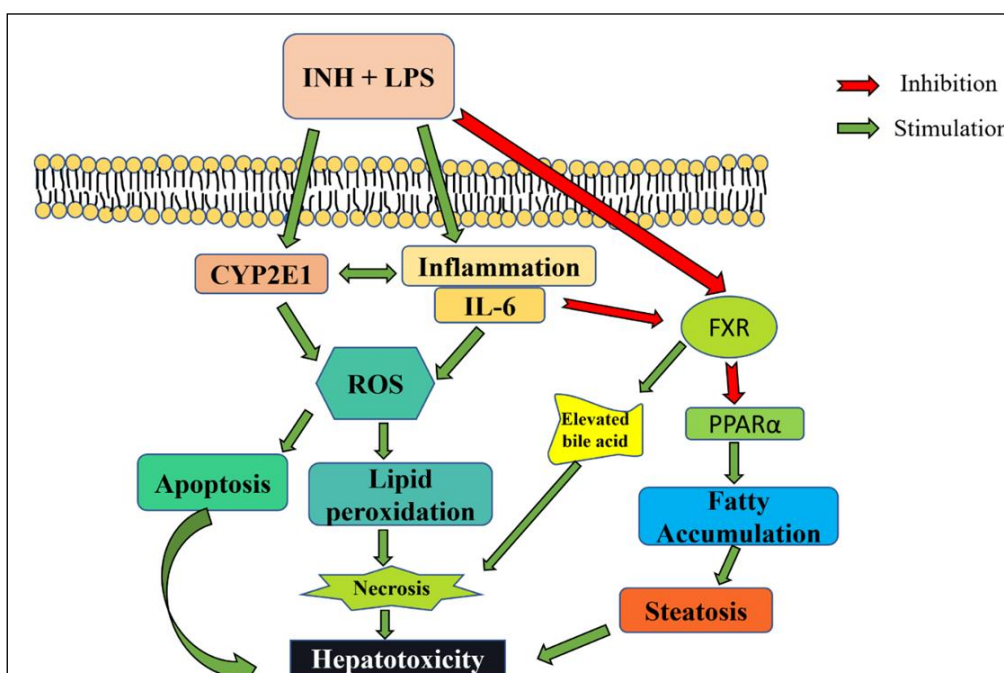


Fig 1: Mechanism of Hepatotoxicity

Liver Function Test

- The following enzymes are included in standard liver function tests (LFTs).
- Alanine Transaminase (ALT)
- Aspartate Transaminase (AST)
- Alkaline Phosphatase (ALP)
- Serum Glutamic-Pyruvic Transaminase (SGPT)
- Serum Glutamic-Oxaloacetic Transaminase (SGOT)
- Gamma Glutamyl Transferase (GGT), Together with
- Bilirubin
- Albumin
- Total Protein and
- Globulin.

Table 1: Hepato protective Plants

| Plant | Common name | Family | Parts used | Hepatotoxicant | Extract used | Bioactive constituents | Evaluate histological parameters |
|-------------------------------------|-------------------------|----------------|--------------------|-----------------------------------|------------------------|---|---|
| <i>Thalictrum Foliolosum</i> [18] | Leafy Meadow-Rue | Ranunculaceae | Root | Paracetamol | Ethanollic | Alkaloids (berberine, jatrorrhizine, palmatine, thalrugosidine, thalrugosaminine, thalisopine thaligosine, thalirugidine, thalirugine, 8-oxyberberine, berlambine | ALT, ASP, AST |
| <i>Ipomoea staphylina</i> Linn [19] | Clustered Morning Glory | Convolvulaceae | Leaves | CCl4 | aqueous extract | Ergotamine, Ergometrine, Ergine, Ergoline, Ergonovine | SGOT, SGPT, ALP and bilirubin. |
| <i>Cucumis ficifolius</i> [20] | Cucumis | Cucurbitaceae | Root | CCl4 | methanol crude extract | | |
| <i>Solanum nigrum</i> [21] | Black nightshade | Solanaceae | Leaves and berries | D-Galactosamine | Hydroalcoholic | Solanine, Sapogenin, Solanidine, Solamargine, Diosgenin | ALT, AST, ALP, |
| <i>Phyllanthus niruri</i> [22] | Stonebreaker | Phyllanthaceae | aerial parts | CCl4 | aqueous extract | Geraniin, Corilagin, Securinine, Lupeol, Astragaline, | AST, ALT, GSH, and SOD levels |
| <i>Dypsis lutescens</i> [23] | Yellow palm | Arecaceae | leaves | D-galactosamine | ethanollic extract | Luteolin, Prechafuroside, orientin, iso orientation | GOT, GPT, ALP, MDA |
| <i>Moringa oleifera</i> [24] | Drumstick | Moringaceae | Bark | Paracetamol | Ethanollic | Quercetin, apigenin, kaempferol, and isorhamnetin | GPT/ALT, GOT/AST, ALP, Total bilirubin, Total protein |
| <i>Prunella vulgaris</i> [25] | Selfheal | Lamiaceae | Floral spikes | Paracetamol | Hydro alcoholic | ursolic acid, β -amyryn, quercetin, α -spinasterol, stigmasterol, β -sitosterol, daucosterol. | SGPT, SGOT, ALP, Total protein, Direct bilirubin |
| Anbara [26] | Dates | Arecaceae | Fruits | CCl4 | Aqueous | Palmitic acid, Syringic acid, Procyanidin, Fructose, β -carotene, p-coumaric acid, ferulic acid, sinapic acids, | AST, ALP, ALT, RBC, WBC, MCV, Hb, |
| <i>Aerva sanguinolenta</i> [27] | Mountain knotgrass | Amaranthaceae | Whole plant | Acetaminophen | Ethanollic | Ferulic acid, Syringic acid, Narcissin, Feruloyltyramine, sitosteryl palmitate, α -amyryn, botulin, β -Sitosterol | GOT, GPT, ALP, LDH |
| <i>Tectona grandis</i> [28] | Teak | Lamiaceae | Bark | CCl4 | Ethanollic | Phthalic acid, n-hexadecanoic acid, 9-Octadecenoic acid | SGPT, SGOT, ALP, Total bilirubin |
| <i>Hippophae rhamnoides</i> [29] | Sea buckthorn | Elaeagnaceae | Leaf | Lead acetate | Aqueous | Vitamin C, Carotenoids, L-ascorbic, Quercetin, rutin acid | AST, ALP, ALT |
| <i>Acacia catechu</i> [30] | Khair | Fabaceae | seed | hydroxyl propyl cellulose | Ethanollic | Quercetin, catechin | AST, ALT, ALP, LDH |
| <i>Bassia Latifolia</i> [31] | Mohua | Sapotaceae | Bark | Carbon Tetrachloride, Alcohol and | Ethanollic | | ALT, ALP, BILD, BILT, ALB, PRO, |

| | | | | Ranitidine | | | CHO, TG, |
|-------------------------------------|--------------------------|---------------|---------------|----------------------------|--|--|---|
| <i>Cordia obliqua</i> [32] | Clammy cherry | Boraginaceae | Fruit | paracetamol | ethanolic and aqueous | Macrophylline, coumarins, β -sitosterol, quercetin, quercitrin | ALT, AST, ALP, direct bilirubin |
| <i>Mallotus repandus</i> [33] | | Euphorbiaceae | Stem | D-Galactosamine | Ethyl Acetate | α -humulene, β -Selinene, Aciphyllene, α -copaen, caryophyllene oxide | ALT, AST, ALP, direct bilirubin, Total protein |
| <i>Dipteracanthus patulus</i> [34] | Spreading Ruellia | Acanthaceae | Whole plant | Paracetamol | Ethyl acetate | Vanilloside, Acteoside, Syringin, Cistanoside E, Isoacteoside | SGOT, SGPT, ALP |
| <i>Schrebera swietenoides</i> [35] | Mala plasu | Oleaceae | Fruit | CCl ₄ | Ethanolic | Sitosterol, Gallic acid, Caffeic acid, Rutin, Apigenin-7-glucoside, Catechin, kaempferol | SGPT, SGOT, ALP and Total bilirubin |
| <i>Barleria Cuspidata</i> [36] | Spiny Barleria | Acanthaceae | Whole plant | CCl ₄ | Methanolic | Barlerinoside, barlerin, acetylbarlerin, lupuloside | AST, ALP, ALT, ACP |
| <i>Baliospermum montanum</i> [37] | Willd Muell Arg | Euphorbiaceae | Root | Ethanol & CCl ₄ | Hydroalcoholic | Axillarenic acid, Baliospermin, Montanin | AST, ALT, Bilirubin |
| <i>Acampe praemorsa</i> [38] | Brittle orchid | Orchidaceae | Aerial parts | Ethanol | Hydro-alcoholic | Phenanthraquinone, Amoeylin, Isoamoeylin, Flavanthrin | ALT, AST, ALP, Total bilirubin, Total protein |
| <i>Gardenia gummifera</i> L.f. [39] | Gummy cape jasmine | Rubeaceae | Fruit | CCl ₄ | Methanolic | Furfural, Maltol, Pantanoic acid, | SGPT, SGOT, ALP, Total protein, Total bilirubin |
| <i>Ehretia laevis</i> Roxb [40] | Dant-Rang | Boraginaceae | Flowers | Paracetamol | Hydro-alcoholic | α and β amyryl, piperazine, Gallic acid, Tannic acid, Rutin, Decanoic acids, Phthalic acid, Phytol, | ASAT, ALAT, ALP, and Total Protein |
| <i>Zanthoxylum armatum</i> [41] | Suterberry | Rutaceae | Rhizome | Paracetamol | Ethyl acetate, Chloroform Methanol | Zanthonitrile, Armatamide, Berberine, Limonene, Linalool | AST, ALT, ALP, Total bilirubin, Total protein |
| <i>Foeniculum vulgare</i> [42] | Sweet fennel | Umbellifers | Seed | Paracetamol | Ethanolic | Anethole, Estragole, Fenchone, Limonene, Phellandrene, Pinene | AST, ALP, ALT, Bilirubin |
| <i>Leucas cephalotes</i> [43] | Head Leucas (Dronpushpi) | Lamiaceae | Whole plant | Paracetamol | Ethanolic | Oleanolic acid, Ursolic acid, 3-sitosterol, α & β Sitosterol, Asperphenamate, Maslinic acid, Linifoliside, Nectandrin B | ALT, AST, ALP, Bilirubin |
| Olive Leaf [44] | <i>Olea europaea</i> | Oleaceae | leaves | CCl ₄ | Super Critical Carbon Dioxide | Oleuropein, Hydroxytyrosol, Luteolin, Rutin, Caffeic acid, Catechin | ALT, AST, ALP, LDH |
| <i>Actiniopteris radiata</i> [45] | Ray fern | Pteridaceae | Root | Paracetamol | Hexane, Ethyl acetate, Hydro-alcoholic | Hentriacontane, Hentriacontanol, β -sitosterol, Quercetin-3-rutinoside (Rutin), β -sitosterol palmitate | SGPT, SGOT, ALT, AST, ALP, Total bilirubin |
| <i>Pentatropis nivalis</i> [46] | White Milkweed | Apocynaceae | Aerial part | CCl ₄ | Methanol | Acokanthera, Apocynum, Cerbera, Nerium | SGPT, SGOT, ALP, Cholesterol, Total bilirubin |
| <i>Bassia latifolia</i> Roxb [47] | Butter Tree | Sapotaceae | Leaves, Fruit | Paracetamol | Ethanolic | Madhucic acid, Farnesol, α -Farnesene, Dendrolasin, 2,3-Dihydrofarnesol, | SGPT, SGOT, SALP, Total bilirubin, Direct bilirubin |
| <i>Hibiscus surattensis</i> [48] | Bush Sorrel | Malvaceae | Aerial parts | CCl ₄ | Ethyl acetate, Methanol | Kaempferol, Morin, Trifolin, Quercetin, Spiraeoside | ALT, AST, ALP, Total protein, Total bilirubin |

| | | | | | | | |
|--|----------------|---------------|--------|-------------|------------|--|--|
| Chenopodium album linn ^[49] | Goosefoot | Amaranthaceae | Leaves | Paracetamol | Methanolic | β -carotene, Catechin, Gallic acid, Caffeic acid, p-Coumaric acid, Ferulic acid, β -sitosterol | SGOT, SGPT, ALP, Direct bilirubin, Total bilirubin |
| Moringa oleifera ^[50] | Drumstick tree | Moringaceae | Leaves | Paracetamol | Nil | Quercetin Astragaloside, Kaempferol, Apegenin, Genistein, Epicatechin | ALT, AST, Gamma Glutamyl Transferase (GGT) |

Conclusion

The numerous pharmacological qualities of therapeutic plants that have been examined experimentally were gathered in this study. However, it is critical to recognise and identify and describe the lead molecule produced from plants that may have the capacity to protect the liver. The hepatoprotective effects of medicinal plants may be due to their ability to reduce oxidative stress and modulate metabolic pathways that leads to hepatotoxicity. Medicinal plants that are high in phytochemicals have high antioxidant activity, which helps to prevent liver damage. These data could be used to justify additional research into the comprehensive pharmacological evaluation of hepatoprotective medicinal plants.

Reference

- Sumaia A, Ali Noha H, Sharief Yahya S, Mohamed. Hepatoprotective activity of some medicinal plants in Sudan. Evidence-Based Complementary and Alternative Medicine. 2019. <https://doi.org/10.1155/2019/2196315>
- Umakrithika S, Manimekalaib P, Kumar R, Senthil, Manna PK, Kannan K. *In-Vitro* Hepatoprotective Activity of Lindernia Madayiparensis extract. 2019. <http://dx.doi.org/10.2139/ssrn.3370122>
- Wang Guo-Kai, Zhang Nan, Wang Yi, Liu Jin-song, Wang Gang, Zhou Zhong-yu *et al.* The hepatoprotective activities of Kalimeris indica ethanol extract against liver injury *in vivo*. Food science & nutrition. 2019; DOI:10.1002/fsn3.1241
- Hassan Towseef, D Veera kumar, Naseer Insha N Anandhi. Hepatoprotective activity of some medicinal plants: a review. International research journal of pharmacy,2019:10(5). DOI: 10.7897/2230-8407.1005154
- Birhanu Geta Meharie, Gedefaw Getnet Amare, Yaschilal Muche Belayneh. Evaluation of hepatoprotective activity of the crude extract and solvent fractions of Clusia abyssinica (Euphorbiaceae) leaf against CCl₄-induced hepatotoxicity in mice. Journal of experimental pharmacology, 2020, 12. DOI <https://doi.org/10.2147/JEP.S248677>.
- Ielciu I, Sevastre B, Olah NK, Turdean A, Chis e E, Marica R *et al.* Evaluation of hepatoprotective activity and oxidative stress reduction of Rosmarinus officinalis L. shoots tincture in rats with experimentally induced hepatotoxicity. Molecules, 2021, 26. Doi: <https://doi.org/10.3390/molecules26061737>
- Shabbir M, Afsar T, Razak S, Almajwal A, Khan MR. Phytochemical analysis and Evaluation of hepatoprotective effect of Maytenus royleanus leaves extract against anti-tuberculosis drug induced liver injury in mice. Lipids in health and disease,2020:19(1). Doi:10.1186/s12944-020-01231-9
- Mohammad K, Parvez Mohammed S, Al-Dosari, Ahmed H, Arbab, Perwez Alam, Mansour S. *et al.* Hepatoprotective effect of Solanum surattense leaf extract against chemical induced oxidative and apoptotic injury in rats. BMC Complementary and Alternative Medicine. 2019, 19. <https://doi.org/10.1186/s12906-019-2553-1>
- Hashem MM, Salama MM, Mohammed FF, Tohamy AF, El Deeb KS. Metabolic profile and hepatoprotective effect of Aeschynomene elaphroxylon (Guill. & Perr.). Plos one,2019:14(1). Doi: <https://doi.org/10.1371/journal.pone.0210576>
- Mohamed Mirghani, Hassan S, Khalid Mohammed Mona S, Mohamed Ahmed Babiker, Ali Amna, Wadah Osman *et al.* Hepatoprotective and free radical scavenging activities of methanol extract fractions of Capparis decidua Edgew (Forssk.) (Capparidaceae). African journal of pharmacy and pharmacology,2020:14(8). Doi: 10.5897/AJPP2020.5180
- Sandhu Naemat. Navarro Victor. Drug-Induced Liver Injury in GI Practice. Hepatology Communications,2020:14(8). Doi: <https://doi.org/10.5897/AJPP2020.5180>
- Teschke Rolf. Hepatotoxicity: molecular mechanisms and pathophysiology. International journal of molecular sciences,2019:20(1). Doi: <https://doi.org/10.3390/ijms20010211>
- Vilas-Boas Vânia, Vinken Mathieu. Hepatotoxicity induced by nanomaterials: mechanisms and *in vitro* models. Arch Toxicol.,2021:95(1). Doi: 10.1007/s00204-020-02940-x.
- Yuvaraja KR, Santhiagu A, Jasemine S, Gopalsathees Kumar K. Hepatoprotective activity of Chloroform and ethyl acetate extract of Dipteracanthus patulus against paracetamol induced hepatotoxicity in rats through antioxidant mechanism. Research journal of pharmacy and technology,2020:13(1). Doi: 10.5958/0974-360X.2020.00041.4
- Rotundo Laura, Pyrsopoulos Nikolaos. Liver injury induced by paracetamol and challenges associated with intentional and unintentional use. World Journal Hepatology,2020:12(4). Doi:10.4254/wjh.v12.i4.125
- Akhtar Md. Shabib, Middha Anil Kumar, Kumar Vinay. Callicarpa macrophylla leaves extracts evaluated as hepatoprotective in wistar rats. Journal of drug delivery and therapeutics,2019:9(3-s). Doi: <http://dx.doi.org/10.22270/jddt.v9i3-s.3087>
- Teschke, Rolf. Hepatotoxicity: molecular mechanisms and pathophysiology. International journal of molecular sciences,2019:20(1). Doi:10.3390/ijms20010211
- Gregory Marslin, Jose Prakash. Hepatoprotective activity of thalictrum foliolosum (ranunculaceae) root ethanolic extract. International journal of life science

- and pharma research,2020:10(3). Doi: <http://dx.doi.org/10.22376/ijpbs/lpr>
19. Jeyadevi Ramachandran, Arul Ananth Devanesan and Sivasudha Thilagar. Hepatoprotective and antioxidant activity of *Ipomoea staphylina* Linn. *Clinical Phytoscience*,2019:5. Doi: <https://doi.org/10.1186/s40816-019-0112-4>
 20. Mebrahtu Arayaa Ephrem, Anteneh Adamuc Betelhem, Periasamyb Gomathi *et al.* *In vivo* hepatoprotective and *In-vitro* radical scavenging activities of *Cucumis ficifolius* a rich root extract. *Journal of Ethnopharmacology*, 2019, 242. Doi: <https://doi.org/10.1016/j.jep.2019.112031>
 21. Chester Karishma, Zahiruddin Sultan, Ahmad Adil, Khan Washim, Paliwal Sarvesh, Ahmad Sayeed. Bioautography-based identification of antioxidant metabolites of *solanum nigrum* L. and exploration its hepatoprotective potential against D-galactosamine-induced hepatic fibrosis in rats. *Pharmacognosy Magazine*, 2019, 15. Doi:10.4103/pm.pm_359_18
 22. Ezzat Marwa I, Okba Mona M, Ahmed Sherif H, El-Banna Hossny A, Prince Abdelbary, Mohamed Shanaz O, *et al.* In-depth hepatoprotective mechanistic study of *Phyllanthus niruri*: *In vitro* and *in vivo* studies and its chemical characterization. *Plos one*,2019:15(1). Doi: <https://doi.org/10.1371/journal.pone.0226185>
 23. El-Ghonemy MM, El-Kashak WA, Mohamed TK, Omara EA, Hussein J, Farrag ARH *et al.* Hepatoprotective activity of *Dypsis lutescens* against D-galactosamine-induced hepatotoxicity in rats and its phytoconstituents. *Asian pacific journal of tropical biomedicine*,2019:9(11). Doi: 10.4103/2221-1691.270979
 24. Islam Rajibul, Alam Md. Jahir. Evaluation of liver protective activity of *moringa oleifera* bark extract in paracetamol induced hepatotoxicity in rats. *Bio Rxiv*. 2019. Doi: <https://doi.org/10.1101/513002>
 25. Ahmad Gazanfar, Masoodi Mubashir H, Tabassum Nahida, Ahmad Mir Sameer. *In vivo* hepatoprotective potential of extracts obtained from floral spikes of *prunella vulgaris* L, *Journal of Ayurveda and Integrative Medicine*. 2019. Doi: <https://doi.org/10.1016/j.jaim.2019.08.003>
 26. Al-Radadi, Najlaa S, Adam Shama IY. Green biosynthesis of pt-nanoparticles from anbara fruits: toxic and protective effects on CCl4 induced hepatotoxicity in wister rats. *Arabian Journal of Chemistry*, 2019. Doi: <https://doi.org/10.1016/j.arabjc.2019.08.008>
 27. Sarker Joy, Ali Md. Rahmat, Khan Muhammad Ali, Rahman Md. Mahbubur, Hossain ASM Sakhawat, Alam AHM Khurshid. The Plant *aerva sanguinolenta*: A review on traditional uses, phytoconstituents and pharmacological activities. *Pharmacog rev*,2019:13(26). Doi: 10.5530/phrev.2019.2.9
 28. Bagali Rajkumar S, Jalalpure Sunil S, Patil SS. *In-vitro* antioxidant and *in-vivo* hepatoprotective activity of ethenolic extract of *tectona grandis* bark against ccl4 induced liver injury in rats. *Pharmacognosy Journal*,2020:12(3). Doi: 10.5530/pj.2020.12.89
 29. Zargar Rizwana, Raghuvanshi Pratiksha, Koul Aditi Lal, Rastogi Ankur, Khajuria Pallavi, Wahid Aafreen *et al.* Hepatoprotective effect of seabuckthorn leaf-extract in lead acetate-intoxicated wistar rats. *Drug and Chemical Toxicology*, 2020. DOI: 10.1080/01480545.2020.1775630
 30. Thangavelua Lakshmi, Balusamyb Sri Renukadevi, Shanmugama Rajeshkumar, Sivanesan Senthilkumar, Devaraja Ezhilarasan, Rajagopalanc Vijayaraghavan *et al.* Evaluation of the sub-acute toxicity of acacia catechu wild seed extract in a wistar albino rat model. *Regulatory Toxicology and Pharmacology*, 2020, 113. Doi: <https://doi.org/10.1016/j.yrtph.2020.104640>
 31. Thimmaraju Manish Kumar, Mondal Prasenjit, Venu Kola, Padmaja Bookya, Babu Gummadi Sridhar, Kumar Rudra Dinesh *et al.* Carbon tetrachloride, alcohol and ranitidine induced hepatotoxicity and its protection by bark extracts of *bassia latifolia* in wister rats. *Journal of herbs, spices & medicinal plants*, 2020. DOI: 10.1080/10496475.2020.1729286
 32. Tharun G, Sivakrishnan S, Sharma JVC. Toxicity assessment, evaluation of antioxidant and hepatoprotective activity on *cordia obliqua* fruit extracts. *Pharmacognosy Journal*,2020:12(5). Doi: 10.5530/pj.2020.12.142
 33. Mondal Milon, Hossain Md. Monir, Hasan Md. Rakib, Tarun Md. Towhidul Islam, Islam Md. Al Foyjul, Choudhuri MSK *et al.* Hepatoprotective and antioxidant capacity of *mallotus repandus* ethyl acetate stem extract against d-galactosamine-induced hepatotoxicity in rats. *ACS omega*, 2020, 5. Doi: <https://dx.doi.org/10.1021/acsomega.9b04189>
 34. Yuvaraja KR, Santhiagu A, Jasemine S, Kumar K Gopalasathees. Hepatoprotective activity of chloroform and ethyl acetate extract of *dipteracanthus patulus* against paracetamol induced hepatotoxicity in rats through antioxidant mechanism. *Research journal of pharmacy and technology*,2020:13(1). Doi:10.5958/0974-360X.2020.00041.4
 35. Bagali Rajkumar S, Jalalpure Sunil S, Patil SS. Evaluation of *schrebera swietenoides roxb.* fruit ethanolic extract for antioxidant and hepatoprotective activity against ccl4 induced liver injury in rats. *Research journal of pharmacy and technology*,2020:13(11). Doi:10.5958/0974-360X.2020.00895.1
 36. Sheeba Tabassum SS, Rajaram C, Kumar S, Nelson R Manohar, Reddy K. Ravindra. Evaluation of hepatoprotective activity of the methanolic extract of *barleria cuspidata* against ccl4 induced liver damage in experimental rats. *Research Journal of Pharmacy and Technology*,2020:13(2). Doi:10.5958/0974-360X.2020.00101.8
 37. Ahirwar B, Ahirwar D. Antioxidant and Hepatoprotective Activity of Root extract of *Baliospermum montanum* (Willd) Muell Arg. *Research Journal of Pharmacy and Technology*,2019:12(6). Doi: 10.5958/0974-360X.2019.00452.9
 38. Uppala PK, Rao GSNK, Reddy AR, Umasankar K, Joshitha C, Anusha K *et al.* Hepatoprotective activity of *acampe praemorsa*. *Journal of pharmaceutical research international*,2021:33(30A). Doi: <https://doi.org/10.9734/jpri/2021/v33i30A31627>
 39. Kumar NM, Vinay, Mahmood Riaz V, Krishna B, Ravishankara Sudhesh L. Shastr. Antioxidant and *in vivo* hepatoprotective effects of *gardenia gummifera* L.f. fruit methanol extract. *Clinical pytoscience*, 2020, 6. Doi: <https://doi.org/10.1186/s40816-020-00188-7>

40. Tarke Santosh Rangnathrao P. Shanmugasundaram. Antioxidant and hepatoprotective activity of *ehretia laevis roxb* against paracetamol induced acute hepatotoxicity in wistar rats. *Research journal of pharmacy and technology*,2019:12(12). Doi: 10.5958/0974-360X.2019.01067.9
41. Talluri MR, Gummadi Veda Priya, Battu GR, Killari KN. Evaluation of hepatoprotective activity of *zanthoxylum armatum* on paracetamol-induced liver toxicity in rats. *Indian journal of pharmaceutical sciences*,2019:81(1). Doi: 10.4172/pharmaceutical-sciences.1000489
42. Nazir Tayyaba, Shakir Lubna, Rahman Zaka-ur, Najam Komal, Choudhary Aqsa, Saeed Nasira *et al.* Hepatoprotective activity of *foeniculum vulgare* against paracetamol induced hepatotoxicity in rabbits. *Journal of Applied Pharmacy*,2020:12(1). Doi: 10.35248/2376-0354.20.12.270
43. Islam G, Gahlot Kavita, Mani Munesh, Kumar Prevesh, Shukla Divaker. Hepatoprotective activity of *leucas cephalotes* against paracetamol induced hepatotoxicity in rats. *Research journal of pharmacy and technology*,2020:13(3). Doi:10.5958/0974-360X.2020.00218.8
44. Taamalli Amani, Anouar Feriani, Jesus Lozano-Sanchez, Lakhdar Ghazouani, Afoua El Mufti, Mohamed S *et al.* Potential hepatoprotective activity of super critical carbon dioxide olive leaf extracts against CCl_4 -induced liver damage. *Foods*,2020:9(6). Doi: <https://doi.org/10.3390/foods9060804>
45. Sankar Kandukuri Gouri, Venkateswarlu Bendi Sri. Phytochemical analysis, antioxidant and hepatoprotective activity of *actinopterin radiata*. *International Journal of Pharmacy and Pharmaceutical Sciences*,2020:12(8). DOI: <http://dx.doi.org/10.22159/ijpps.2020v12i8.37770>.
46. Babre Nilesh P, Gouda T. Shivaraj Gowrishankar Narayanswamy Lachmanan. Hepatoprotective activity of *against Carbon tetrachloride* induced hepatic damage *Pentatropis nivalis* in rats. *Asian journal of pharmacy and pharmacology*,2020:6(6). Doi: <https://doi.org/10.31024/ajpp.2020.6.6.3>
47. Tripathi Deepti, Prof. Dr. Trilochana Yeddu. Combined hepatoprotective effect of leaves and flowers of *bassia latifolia roxb* in paracetamol hepatotoxic rats. *Indian journal of research in pharmacy and biotechnology*,2019:7(4). Doi: <https://doi.org/10.31426/ijrpb I>
48. John Anoop LN, Kannappan P Manojkumar. Evaluation of hepatoprotective activity in methanolic extract of aerial parts of *hibiscus surattensis*. *Research journal of pharmacy and technology*,2020:13(10). Doi: 10.5958/0974-360X.2020.00816.1
49. Das Angana, Borthakur Mridul Kumar. Hepatoprotective activity of *chenopodium album linn.* against paracetamol induced liver damage in albino rats. *International journal of pharmaceutical sciences and research*. 2020:11(11). Doi: 10.13040/IJPSR.0975-8232.11(11).5605-10
50. Ranganathan V, Punniamurthy N, Ahamad D. Basheer, Kumar S, Sathesh. Evaluation of Hepatoprotective activity of *Moringa oleifera* in chicken. *The Journal of Phytopharmacology*,2020:9(3). Doi: 10.31254/phyto.2020.9304